This Page Is Inserted by IFW Operations and is not a part of the Official Record

BEST AVAILABLE IMAGES

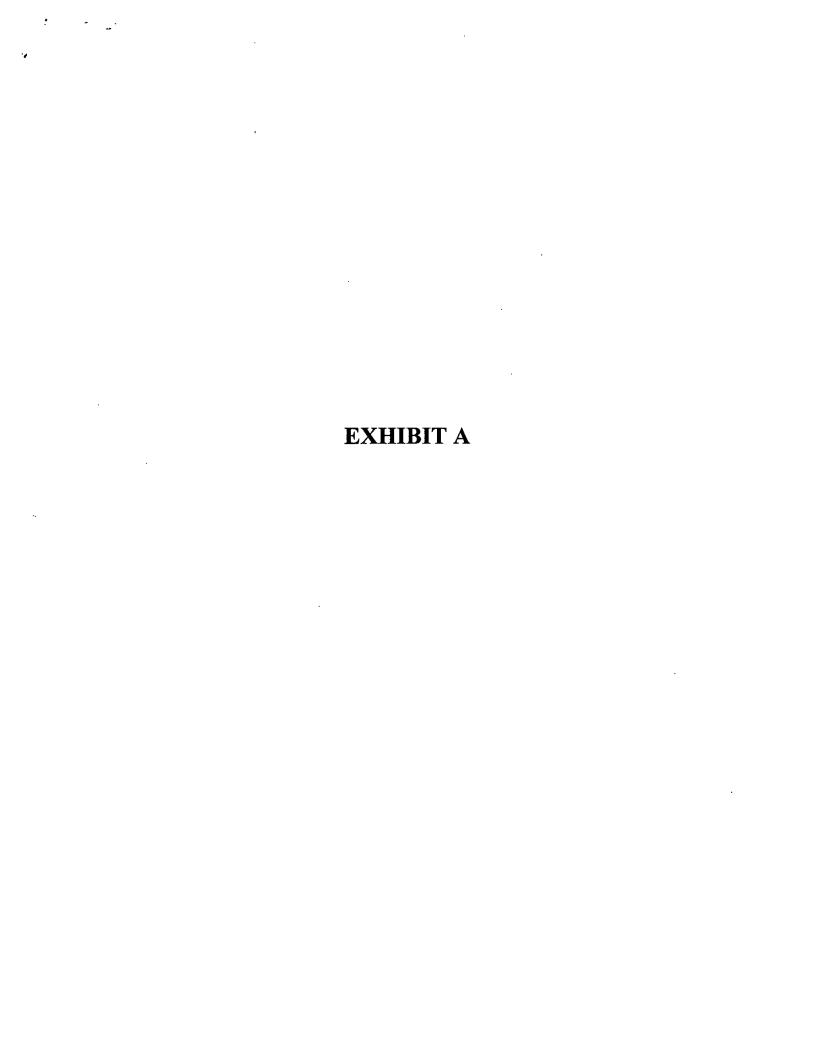
Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

IMAGES ARE BEST AVAILABLE COPY.

As rescanning documents will not correct images, please do not report the images to the Image Problem Mailbox.



ANTIPROLIFERATIVE SYNTHETIC PEPTIDE 2438, INTERFERON ALPHA-2, AND IMMUNOSUPPRESSANT CYCLOSPORINE A. EARLY LYMPHOCYTE ACTIVATION EVENTS ARE INHIBITED BY × FRAGMENT OF HUMAN

Kirill V. Freze*, Alla V. Danilkovich*, Alexey I. Kharitonenkov⁸, Alexander F. Shevalier*, Alexander M. Surin*, Alla B. Khodorova*, Eugene I. Astashkin*, Tamara V. Bulargina*.

Eugene I. Astashkin*, Tamara V. Bulargina*.

Department of Cell Physiology and Immunology, Biological Faculty, Moscow State University, Moscow, Russia;

Department of Bioorganic Chemistry, Biological Faculty,

Moscow State University, Moscow, Russia;

Institute of Biotechnology, Moscow, Russia; Institute of Bioorganic Chemistry, Moscow, Russia.

Received after revision, June 5, 1994 Received April 15, 1994

interferon alpha-2 amino acid sequence 124-138 inhibits proliferation of T-lymphocytes in vitro. Time-course experiments suggest that peptide 2438 affects early stages of lymphocyte activation. Molecular mechanisms of peptide 2438 action were mitogen-stimulated lymphocytes was studied. By western-blotting with monoclonal antibodies against phosphotyrosine peptide 2438 was shown to decrease the phosphotyrosine content of an endogenous protein substrate (M.M.=36 kDa) in human lymphocytes activated with concanavalin A (ConA). Similar effect on tyrosine-specific phosphorylation in the ConA-induced calcium influx in lymphocytes. chelator Fura-2. in human lymphocytes were measured using a fluorescent calcium chelator Fura-2. In contrast to CsA, peptide 2438 did not affect interferon or Cyclosporine A (CsA). Calcium fluxes induced by ConA A synthetic peptide (designated 2438) corresponding to the human sterferon alpha-2 amino acid sequence 124-138 inhibits observed with calcium native

which prevents the clonal expansion. This kind usually accompanied by a decreased expression of genes coding for The main feature of immunosuppressive agents is their ability to transition their receptors. of, resting lymphocytes to proliferation Earlier we reported that the of, inhibition

Cyclosporine A; IFN, interferon; PBS, phosphate buffered saline PBMC, peripheral blood mononuclear cells; pp36, phosphoprotein with appropriate molecular mass 36 kD; P-Tyr, phosphotyrosine; [Ca $^{2+}$], concentration of cytosolic free calcium. The abbreviations used are: ConA, Concanavalin

Physiology and Immunology, Biological Faculty, Moscow State University, 119899, Moscow, Russia Corresponding author: Kirill V. Freze, Department of Cell

Vol. 33, No. 5, 1994

was measured in time-course experiments lymphocyte activation, investigate phosphoproteins extracted from peripheral blood mononuclear cells polyclonal mitogen, and that of CsA. peptide 2438, elucidate the direct effects of antiproliferative agents, of tyrosine phosphorylation. mechanisms of the peptide 2438 action remain unclear. We activity of interferon alpha-2, molecular mass of 36 kDa [2]. In contrast to the original molecule increase in phosphotyrosine content of interferon alpha-2 inhibited mitogen-stimulated synthetic antiproliferative activity may be coupled with the inhibition and CsA on ConA-induced T-cells [1] and downregulated the activation-dependent 2438 with [3] nor affects effects peptide and CsA. To this end, on the processes stimulated ConA and/or antiproliferative agents, 2438 of, peptide 2438 neither possesses inhibition of human We also examined the influence of peptide peptide to compare the peptide 2438 action with from the we determined the tyrosine content lymphocyte viability [1]. Molecular In this study we made an attempt calcium fluxes. 2438 and CsA on early stages of carboxy ø phosphoprotein with a in T-cell lymphocytes by a terminus proliferation In order proliferation synthetic antiviral ဋ CsA and suppose human ဋ

Materials and Methods.

from Amersham. Other chemicals were of reagent grade. (Fura-2/AM) was from "Calbiochem" (Switzerland). Concanavalin A was from Pharmacia, Sweden. Monoclonal antibodies against P-Tyr were described earlier [4]. Radioactive 3H-thymidine was purchased from Sandoz (Sandimmune). Stock solution of CsA (1 mg/ml) was made in dimethyl sulphoxide. Fluorescent calcium chelator Fura-2 G.Chipens (Institute of Organic Chemistry , Riga, Latvia). CsA was All media and media supplements for cell cultures were obtained rom Sigma. Recombinant human IFN (Reaferon) was a gift of Dr.

Separation of cells.

Human blood was obtained from healthy volunteers. Monor cells were isolated by gradient centrifugation according to Mononuclear

Lymphocyte proliferation.

Cells were placed into 96-well tissue culture plates (1x10 5 cell/200 μ l/well). Peptide 2438 (2 μ g/ml) and CsA (0.1 μ g/ml) were added to cells either 15 min before, or simultaneously, or else at

experiments were done in triplicates. Cells were maintained in an atmosphere of 5% CO_2 , 95% relative humidity, at 37°C for 72 hours. H-thymidine (0.5 μ Ci/well) was added 15 hrs before termination of cultivation. The cells were harvested onto glass fiber filters and scintillation counter. the in per cents according to the equation: different radioactivity uptake was determined on an LKB-Wallac beta periods of Inhibition of proliferation was time after the addition of expressed

Per cent of inhibition = experiment (cpm) × 100

where "experiment" stands for an average radioactivity value the presence of antiproliferative agents, whereas "control" is average radioactivity in their absence. Standard deviation was 8. less that 10% in all experiments. The total number of experiments control (cpm) whereas "control" is

<u>Detection of phosphotyrosine in cellular phosphoproteins.</u>

reaction was terminated the cells were centrifuged, the precipitates resuspended in 100 µl of sample buffer for electrophoresis (0.2 % SDS, 10% glycerol, 50 mM dithiothreitol, 62 mM Tris-HCl, pH 6.8), and immediately heated in boiling water for 5 min. Samples (25 µl) were applied to 10% polyacrylamide gels and electrophoresed under denaturing conditions according to Laemmli [6]. Separated proteins were blotted onto nitrocellulose filter using a semi-dry transfer system [7]. To prevent nonspecific binding of antibodies to nitrocellulose, blots were saturated by preincubation in blocking solution (1 % bovine serum albumin in 0.014 M NaCl, 0.005 M Na phosphate, pH=7.4, containing 0.05% of nonionic detergent Tween-20) for 1 hr. The preincubated blots were saturated by the serum of the serum albumin in 0.014 M NaCl, 0.005 M Na phosphate, pH=7.4. stained with monoclonal antibodies against P-Tyr, followed by rabbit antimouse antibodies, conjugated with horseradish peroxidase [4]. All dilutions of the antibodies were made in the blocking solution. The color reaction was developed by a peroxidase substrate, 1-chloro-4-naphtol. The gel lanes containing stained bands of polypeptides were scanned on an Ultroscan laser densitometer, and the optical density values were compared. The 30 min at 37° C. The reaction was started by addition of peptide 2438 (2 μ g/ml), CsA (1 μ g/ml), IFN (1000 U) and/or ConA (2 μ g/ml) to a final volume of 100 μ l. Non-treated cells served as a control. Incubation time varied from 1 to 15 min. After the experiments were repeated 7 times. Eppendorf tubes in aliquotes of 10⁶ the tubes was resuspended in PBS, Freshly separated cells was washed in PBS and distributed and allowed to equilibrate cells per tube. The content of

Measurement of. Ca2+ cytoplasmic concentrations in PBMC

solution without phenol red, All experiments were run Fura-2/AM (acetoxymethyl ether of calcium chelator Fura-2. In brief, immediately afte separation PBMC suspensions were stained by incubating with incubation the cells were This method was earlier detailed by Grynkievicz [8] washed in the same media as above. buffered with 10 mM HEPES (pH 7.35). twice Fura-2) for and resuspended 1 hr in Hank's using after at After

BIOCHEMISTRY and MOLECULAR BIOLOGY INTERNATIONAL

Vol. 33, No. 5, 1994

concentration of $1x10^6$ cells/ml. All measurements of $[Ca^{2^+}]_1$ were carried out at 37° C in quartz cuvettes on an F-4000 Hitachi spectrofluorimeter under stirring, the excitation and emission wavelengths being 336 and 510 nm, respectively. Graphic representation of $[Ca^{2^+}]_1$ was made according to the equation: $[Ca^{2^+}]_1 = 224 \times (R_{0\,b\,s\,e\,r\,v\,e\,d}^{-R_{0\,1}\,n})/(R_{0\,a\,x}^{-R_{0\,b\,s\,e\,r\,v\,e\,d}})$ [8]. The number of experiments totalled six.

Results.

ConA induced proliferation.

simultaneously with the mitogen. Meanwhile, the antiproliferative obtained treated by ConA longer than for effect of peptide 2438 was considerably inhibition 2438 or CsA were added 15 min before the addition of ConA, and the The maximum inhibition of proliferation was achieved when Fig. 1 demonstrates and for cells treated with IFN. SPA completely weaker the results of proliferation experiments. disappeared once t.he 60 in the nin. hoth In contrast, lower Similar agents case when the cells results CsA inhibited added were peptide added were were

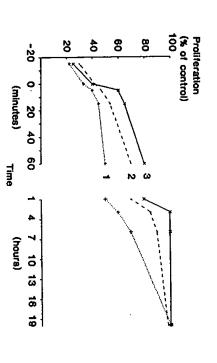


Figure 1.

Human PBMC proliferation in response to ConA in the presence of CsA (1), IFN alpha-2 (2), and synthetic peptide 2438 (3). Drugs were added to PBMC cultures at different time periods before and after the addition of ConA. X-axis is the time of drugs addition, where zero point corresponds to simultaneous addition of ConA and inhibitors of proliferation. Y-axis is proliferation in % in respect to control, where control is proliferation in the absence of drugs.

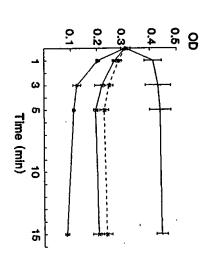


Figure 2.

Phosphotyrosine content of a 36-kDa polypeptide. Y-axis is the optical density values of stained bands ± CEM. X-axis is the incubation time. Cells were incubated with (1) - ConA; (2) ConA + peptide 2438; (3) - ConA + IFN; (4) - ConA + CsA.

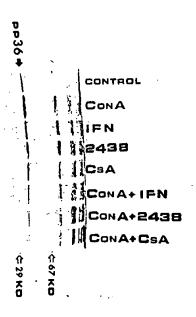
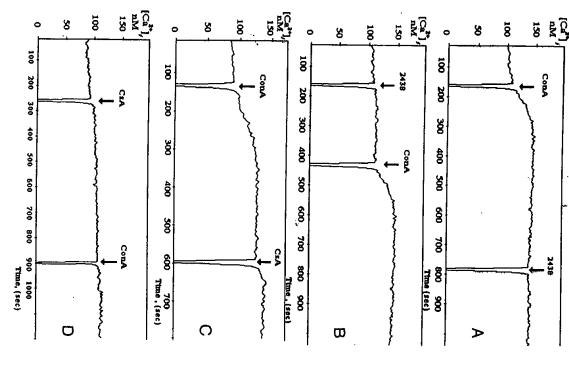


Figure 3.

Western blot of phosphoproteins from human PBMC incubated for 15 min with and without ConA, CsA, IFN and peptide 2438. Non-treated cells serve as a control.



Figures 4 A, B, C, D.

Concentration of cytosolic free calcium in PBMC under treatment with ConA, CsA and peptide 2438. X-axis is the time in minutes. The time and sequence of reagents added are indicated by arrows. Y-axis is Ca^{2} concentration in nanomoles.

proliferation when added long after ConA, and this effect was significant even after 6 hrs of cell cultivation in the presence of the latter (data not shown).

Protein tyrosine phosphorylation.

only in the ConA-treated cells. antiproliferative agents induced a decrease in the P-Tyr polypeptide (pp36) with the maximum P-Tyr level reached in 3 antiproliferative agents alone remained unchanged IFN or CsA (Fig 2). level) (Fig. 3). ConA rapidly induced phosphorylation of a tyrosine phosphorylation. treatment with ConA as well as effects of peptide 2438 and CsA on of a Fig. 2 shows time-dependent changes in pp36 36-kDa to the cell culture markedly decreased the P-Tyr levels ç Simultaneous addition of ConA along with peptide 2438, phosphoprotein values below The P-Tyr content in cells treated with extracted from PBMC after the cells the control phosphotyrosine (Fig. (at QJ content content contro] 36-kDa min

Cytosolic Ca2+ concentration.

(Fig in PBMC. 2438 and CsA differently influenced ConA stimulated calcium fluxes increments of $\left\{\operatorname{Ca}^{2^+}\right\}_1$ in all experiments (Fig 5 B). (Fig 4 B, 5 B). In the meantime, CsA entirely blocked ConA-induced concentrations of 5-10 nM both when added before and peptide 2438 did not prevent the mitogen-induced ${ t [Ca^{2^+}]}_1$ elevation peptide was added before or after ConA (Fig 4 A, was almost independent of peptide at 140 nM, the cells resulted in $[Ca^{2+}]_1$ increase, non-activated cells was about 100 nM. The concentration of cytosolic free calcium in the cytoplasm **≯** whereupon plateaued (Fig 4 A,B). CsA, as it is, 2438, no increased Addition of the mitogen to which reached its maximum matter whether the In contrast, [Ca2+]1 $[ca^{\xi+}]_1$ 5 A). Thus, peptide after ConA values at Moreover, of f

Discussion.

In the preceding papers we studied the activities of various synthetic peptides derived from the carboxy terminal part of the human interferon α -2 molecule [1,2,9]. Only one of them, namely

í

Vol. 33, No. 5, 1994

2438, demonstrated antiproliferative activity against T-lymphocytes stimulated by ConA.

antiproliferative action mechanisms for IFN and peptide 2438. be indicative of at least partial resemblance required conditions similar to those for peptide 2438, stages of lymphocyte activation. The time-course experiments also observed provided the revealed that manifestation of IFN antiproliferative activity strongly suggest that peptide 2438 affects preferentially early after the addition of ConA. Taken together, these findings preferentially on non-stimulated cells and interferes with T-cell from the cell surface. It seems likely that peptide 2438 acts peptide 2438 cannot be explained by the loss of peptide receptors capacity [9], the peptide was added to activated cells after 24 hrs of cultivation inhibition of ConA-induced proliferation by peptide activation. [1]. Since the activated cells retained their peptide binding when added together with a mitogen, being undetectable once the The antiproliferative effect of peptide 2438 was manifested only The time-course resistance to the antiproliferative action of peptide was added before or immediately experiments clearly show that which may 2438 of, Was

Our data with regard to Csa confirmed the result presented earlier by Lin that the inhibition of proliferation was severely diminished after 6 hrs of incubation with a polyclonal stimulator [10].

Recent observations demonstrated the key role of protein tyrosine phosphorylation in lymphocytes activation. T-cells stimulation by polyclonal activators leads to increase in tyrosine phosphorylation of endogenous substrates of protein tyrosine kinases, in particular the ξ chain of CD3 [11], and several polypeptides with molecular masses of 120, 80, 40 kDa [12]; 110, 90, 80 kDa [13]; 100, 84, 57, 38 kDa [14]; 94-100, 90, 64-75, 50-55, 36-40, 21 kDa [15].

Earlier we reported that peptide 2438 was able to lower P-Tyr content in the 36 kDa phosphoprotein (pp36), though measured 1 hr after the cells treatment with the peptide [2]. In this connection it was of interest to study the time course of ConA-induced tyrosine phosphorylation in the presence of peptide 2438, IFN and the widely used immunosuppressant CsA. The data obtained in this

study agree with the previous observations that pp36 phosphorylation is a marker of lymphocyte activation. The kinetic data presented here enable us to suppose that pp36 tyrosine phosphorylation is associated with early activation events.

It is generally accepted that antiproliferative activity of CSA also involves the inhibition of early stages of T-cell activation [10,16]. However, despite recent extensive investigations, the exact molecular mechanisms of CsA action are still unknown. Also, little is known on direct effects of CsA on a cascade of molecular events which follow the antigen-receptor binding.

sensitive hematopoietic cell types [17]. the GO/G1 phase of the cell cycle accompanied by a significant antiproliferative action. antiproliferative action of CsA and IFN. The latter were earlier phosphorylation reduction in the levels of c-myc oncogene mRNA in a reported substrate In particular, they decreased the P-Tyr content in the endogenous studied The present investigation (CsA, ç pp36, have common in PBMC treated with the polyclonal mitogen ConA. IFN and which can be a common constituent Both agents caused a growth peptide demonstrates that all the agents elements 2438) affect in their modes variety arrest tyrosine of of

action of peptide 2438 may of course differ from that of IFN, phosphorylation in our experimental model was similar to have much in common. the features of their action (as well as for IFN and CsA) seem remains to be answered. The mechanism of the interferon molecule possesses the antiproliferative action of IFN is still obscure. both CsA and IFN. The role of the amino acid stretch 124- 138 in The action of question of whether or not this fragment of the synthetic antiproliferative activity yet peptide 2438 on antiproliferative The most tyrosine

We have obtained no direct evidence of the involvement of pp36 tyrosine phosphorylation in the cell growth regulation. It is only a good correlation between modulations of pp36 tyrosine phosphorylation and proliferation of cells that has been noted. Additional experiments would be essential for analysing the role of pp36 in the regulation of lymphocyte proliferation.

Vol. 33, No. 5, 1994

T-cell line HUT-78 [23]. which correlates with the data reported by Vereb for a human Moreover, in our experiments, CsA itself slightly induces $[Ca^{2+}]_1$, confirm the previous observations that CsA inhibits increase obtained at times of CsA preincubation less than 30 min and the alternative interpretation of his data, the more so that they were close examination of the results of Metcalf [19] suggests an cytoplasmic calcium concentration and mitogen-induced calcium inhibit $[Ca^{2+}]_1$ increase [19,20]. More recently, influx are controversial, T-cell activation [10,18], but the data on direct CSA influence on CSA is believed speculations. Some reports demonstrated that CsA did not activation-associated to suppress the cytosolic $[Ca^{2+}]_1$ elevation [21]. However, lower င် inhibit calcium dependent pathways of the than 10 although this problem is a subject cytosolic $[Ca^{2+}]_1$ elevation [22]. $\mu g/m1$. Other recent results CsA has been 'n

further studies are needed to finally elucidate the mechanisms activation nor results). All these findings agree with an assumption that peptide should be noted that peptide 2438 did not affect the production of cytosolic calcium rise is sufficient for IL-2 production [24]. It affected neither mitogen induced nor basal $[Ca^{2+}]_1$ levels in the peptide 2438 action. IL-2 in ConA-activated human Another finding is that, of non-activated cells. not affect Ca-dependent it interferes with PBMC (A.V.Danilkovich, in contrast According to the current view, IL-2 production. pathways ç CsA, peptide 2438 unpublished lymphocyte However, o f

The authors wish .Glotov (Institute of Molecular Genetics, Moscow) iscussion and critical reviewing of the manuscript. ç express their appreciation for to Dr. valuable Boris

Danilkovich, A.V., Freze, K.V., Shevalier, A.F., Samukov, V.V., Kirkin, A.F., and Gusev, MV. (1992) Immunol. Lett. 31, 15-20. Danilkovich, A.V., Kharitonenkov, A.I., Freze, K.V., Shevalier A.F., Kolosova, O.V., Bulargina, T.V., Kirkin, A.F., and Gusev M:V. (1992) FEBS Lett. 296, 271-273

- Shevalier, A.F., Samukov, V.V., Ofitserov, V.I., Kalashnikov,V., Mizenko, G.A., and Kolokoltsov, A.A. (1990) Bioorg. Khim. (USSR) 16, 916-925.
- Kharitonenkov, A.I., and Bulargina, T.V. (1991) Biokhimia -(USSR) 56, 236-242.
- 765 Boyum, A. (1968) Scand. J. Clin. Invest. 21, 97-97. Laemmli, U.K. (1970) Nature 227, 680-685. Kynse-Andersen, A. (1984) J.Biophys. Biochem. Meth. 10, 203-209.
- 8 Grynkievicz, G.M., Poenie, M., and Tsien, R.Y. (1985) J.Biol.
- Danilkovich, A.V., Kharitomenkov, A.I., Freze, K.V., Kolosova, O.V., Bulargina, T.V., Kirkin, A.F., and Gusev, M.V., (1991) FEBS Lett. 295, 70-72. FEBS Lett. 295, Chem. 260, 3440-3440.
- 10. Lin, C.S., Boltz, R.C., Siekierka, J.J., and Sigal, N.H.

- (1991) Cell. Immunol. 133, 269-284.

 11. Samelson, L.E., Patel, M.D., Weissman, A.M., Harford, J.B., and Klausner, R.D. (1986) Cell 46, 1083.

 12. Schwinzer, R., Franklin, R.A., Domenico, J., Renz, H., and Gelfand, E.W. (1992) J. Immunol. 148, 1322-1328.

 13.Alberola-ila, J., Places, L., Cantrell, D.A., Vives, J., and Lozano, F. (1992) J. Immunol. 148:1287-1293.
- Saltzman, E.M., Thom, R.R., and Casnellie, J.E. (1988) J. Biol. Chem. 263, 6956-6959.
- 15. Boyer, S., Ley, S., Davies, A., Ley, S., Davies, A., Ley, S., Ley and Crumpton (1993) Mol.
- Transplantation 51, 1276-1282. Yonishrouach, E., Kimchi, A., Rubinstein, M., (1991)
- 18. Mattila, P.S., Ullman, K.S., Fiering, S., Emmel, E.A., McCutcheon, M., Crabtree, G.R., and Herzenberg, L.A. (1990) EMBO J. 9, 4425-4433.
- 19. Metcalf, S. (1984) Transplantation 38, 161-161.
- Redelman, D. (1988) Cytometry 9, 156-156. Gelfand, E.W., Cheung, R.K., and Mills, G.B. (1987) J.
- Immunol. 138, 1115-1115.
- Tsao, P.W., Diaz, R.J., Radde, I.C., Wong, P.Y., Martell, M.F., Line, J.M., Wilson, G.J., and Coles, J.G. (1991)
 Transplantation 23, 345-349.
 Vereb, G., Panyi, G., Balazs, M., Matyus, L., Matko, J., and Dajanovich, S. (1990) BBA 1019, 159-165.
 Mills, G.B., Cheung, R.K., Grinstein, S., and Gelfand, E.W. (1985) J.Immunol. 134, 1640-1640.

CONTENTS

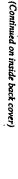
Volume 33, Number 4, July 1994

Biochemistry

Molecular Biology International

a rapid communication journal

Pages 817-1032 Volume 33, Number 5, August 1994 ISSN 1039-9712



743

